Policies and Procedures for

NW 615MUH & NW 636 MUH

Surgical Field Set-up:

Turn on microbead sterilizer for instrument sterilization between animals before you start so it will heat up to sterilization temperature <300°C

Put blue pad down onto heating pad which is on surgical bench

Use sterile gloves to:

Place sterile field drape next to blue pad for instruments and gauze

Open all needed instruments and place onto sterile pad

Place Sterile gauze for surgical procedure and for keeping animal warm during

experiment onto sterile field – made by surgeon day before –

Place Sterile Q-tips (cotton tip applicators) next to surgical area – made by surgeon day before

Place Sterile suture next to surgical area – made by surgeon day before

Fill stainless steel bowl with 70% alcohol for instrument cleaning between animals

Anesthetize animal

Place animal onto metal board and secure using loose loop tape technique

Shave surgical area

Use sterile cotton-tip applicators or gauze to sterilize surgical area with betadine and 70% alcohol – repeat three times.

Use sterile gloves during surgery.

Perform surgery

Between animals all instruments must be:

Washed in 70% alcohol

put into microbead sterilizer for ~ 10-30 seconds

put into 70% alcohol to cool and sterilize

placed onto sterile field drape to dry (be sure not to put instruments into animal with alcohol on them as it may induce tissue injury)

 \*\*Instruments can be used on a maximum of five animals before it is required to be re-steam or gas sterilized – with bead sterilization used between animals.

Surgical Etiquette:

-Always ensure your animals are properly anesthetized: They don’t respond to whisker touch and toe pinch reflex.

-Shave and sterilize appropriately (see above) surgical areas.

-perform surgery according to procedure outlined in IACUC protocol.

-If opening multiple layers (skin and muscle) these layers must be sutured separately.

Vicryl for muscle layer –can be sutured in continuous fashion

PDS for skin layer – must be sutured in interrupted fashion

-When surgery is completed the animal can be returned to his cage. This cage must be kept half on a heating pad.

-You must remain with your animals at all times while they are under anesthesia. If you have to leave the room you must ask someone to sit with the animals in your place.

-Once the animals begin to right themselves (come out of anesthesia) you must administer pain management, ie. Burpenorphine 0.1cc. Record this on the log sheet NW 636 MUH

\*\* All anesthesia taken must be recorded on the appropriate log sheet. See Lauryn or Aaron for directions on this. This is a federally regulated, ie., DEA regulation and will result in Dr. Billiar losing his controlled substance license if this isn’t done correctly.

\*\* Aaron is in charge of all controlled substance dilutions. If you need more of anything, come down to NW 636 MUH and see him.

\*\* Please clean up after you are finished with your surgery. Wash all instruments and autoclave them. Wipe down surgical area and dispose of all sharps into sharps container (DO NOT RECAP NEEDLES)

\*If the biohazard bag is full, tie it up and place it into the large biohazard box

\*\* When you are finished with your animal cage, dump the bedding into a biohazard bag and rinse out the cage. Place the dirty cage with the wire top and filter top next to the sink.

\*\* All animals must have food and water available when housed for any length of time.

\*\* Be sure to fill out surgical (blue) procedure card and place on cage post op. Be sure to include pain management dose and V.

\*\* Be sure to fill out the overnight log sheets for animals who will be housed in our surgical suites for extended lengths of time.